RESEARCH ARTICLE



Antiprotozoal and antimycobacterial activities of *Persea americana* seeds

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Abstract

Background: *Persea americana* seeds are widely used in traditional Mexican medicine to treat rheumatism, asthma, infectious processes as well as diarrhea and dysentery caused by intestinal parasites.

Methods: The chloroformic and ethanolic extracts of *P. americana* seeds were prepared by maceration and their amoebicidal, giardicidal and trichomonicidal activity was evaluated. These extracts were also tested against *Mycobacterium tuberculosis* H37Rv, four mono-resistant and two multidrug resistant strains of *M. tuberculosis* as well as five non tuberculosis mycobacterium strains by MABA assay.

Results: The chloroformic and ethanolic extracts of *P. americana* seeds showed significant activity against *E. histolytica, G. lamblia* and *T. vaginalis* ($IC_{50} < 0.634 \mu g/ml$). The chloroformic extract inhibited the growth of *M. tuberculosis* H37Rv, *M. tuberculosis* MDR SIN 4 isolate, three *M. tuberculosis* H37Rv mono-resistant reference strains and four non tuberculosis mycobacteria (*M. fortuitum, M. avium, M. smegmatis* and *M. absessus*) showing MIC values \leq 50 µg/ml. Contrariwise, the ethanolic extract affected only the growth of two mono-resistant strains of *M. tuberculosis* H37Rv and *M. smegmatis* (MIC \leq 50 µg/ml).

Conclusions: The CHCl₃ and EtOH seed extracts from *P. americana* showed amoebicidal and giardicidal activity. Importantly, the CHCl₃ extract inhibited the growth of a MDR *M. tuberculosis* isolate and three out of four monoresistant reference strains of *M. tuberculosis* H37Rv, showing a MIC = 50 μ g/ml. This extract was also active against the NTM strains, *M. fortuitum, M. avium, M. smegmatis* and *M. abscessus*, with MIC values <50 μ g/ml.

Keywords: Medicinal plant, Seeds, Antimycobacterial, Antiprotozoal, Persea americana

Background

Persea americana Mill. (Lauraceae) is an edible fruit commonly known as *aguacate* (avocado) that grows throughout the tropics. The seeds (crude or toasted) are employed in traditional Mexican medicine to treat skin rashes, diarrhea, and dysentery caused by helminths and amoebas, for the cure of infectious processes caused by fungi and bacteria, as well as for the treatment of asthma, high blood pressure, and rheumatism [1-5]. The seeds of *P. americana* used alone or mixed with other species, such as *Psidium guajava, Mentha piperita* or *Ocimum basilicum*, are mainly employed for the treatment of diarrhea [4].

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Interestingly, the hypolipemic effect of the MeOH extract obtained from *P. americana* seeds has been demonstrated in male rats with induced hypercholesterolemia



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[10,11]. This extract reduced total cholesterol levels, triglycerides and Low density lipoprotein (LDL); on the other hand, increased the levels of High density lipoprotein (HDL). The same effect was described for the aqueous extract, which also reduced blood pressure both in normal rats and those with high blood pressure; in addition, it exerted a hypoglycemic effect on rats and rabbits with diabetes [12-15]. The aqueous extract showed a median Lethal dose (LD₅₀) = 10 g/kg in rats when it was administered orally. Importantly, it did not alter the hematological parameters nor the levels of Alanine aminotransferase (ALT), Aspartate aminotransferase (AST), albumin, and creatinine in male and female rats were treated for 28 days [16].

The hexanic and MeOH seed extracts of P. americana have been described to have a Minimum inhibitory concentration (MIC) of <1.25 µg/ml against Candida ssp., Cryptococcus neoformans and Malassezia pachydermatis. These extracts were also active against Artemia salina, with Lethal concentration (LC50) values of 2.37 and 24.13 mg/ml, respectively. They were also active against Aedes aegypti larvae with LC₅₀ values of 16.7 and 8.9 mg/ml, respectively [17]. On the other hand, the MeOH extract from P. americana leaves inhibited completely the growth of M. tuberculosis H37Ra $(MIC = 125 \ \mu g/ml)$ and H37Rv $(MIC = 62.5 \ \mu g/ml)$; furthermore, the hexane fraction inhibited the growth of both mycobacteria with MIC = $31.2 \ \mu g/ml$ [18]. In addition, the EtOH extract was active against both Gram-positive and -negative bacteria (with the exception of Staphylococcus epidermis and Escherichia coli) with MIC = 500 μ g/ml [19]. Regarding the bacterial activity of P. americana (var Hass and Fuerte), the acetone seed extract exhibited moderate activity against Bacillus cereus, Staphylococcus aureus and Listeria monocytogenes [9]. The trypanomicidal activity of the MeOH extract from *P. americana* seeds has been also tested [20]. It showed moderate activity when was evaluated at the concentration range of 250–500 μ g/ml. In the case of the aqueous seed extract, it had a slight anti-Giardia duodenalis (syn G. lamblia) activity, inducing 23% of mortality at 4 mg/ml [21].

An important antioxidant activity (AOA) of the MeOH extract of *P. americana* seeds and leaves has been described by different methods [8,22-24]. Besides, AOA has been reported in the 100% EtOAc, 70% acetone and 70% MeOH of the peel, pulp and seed extract [9].

Up to now, the activity of the chloroformic (CHCl₃) and ethanol (EtOH) extracts obtained from *P. americana* seeds against anaerobic protozoan and *M. tuberculosis* H37Rv strains with different level of drug resistance has not yet been evaluated. Therefore, herein, the activity of both extracts was tested against the anaerobic protozoa *Giardia lamblia, Entamoeba histolytica* and *Trichomonas* *vaginalis.* In addition, their antimycobacterial activity was evaluated against four mono-resistant reference strains of *M. tuberculosis* H37Rv, two MDR *M. tuberculosis* clinical isolates and five non-tuberculosis mycobacterium (NTM).

Methods

Plant material

P. americana seeds were obtained from the town of Ario de Rosales in the state of Michoacan, Mexico in August 2009. This material was then dried at room temperature and under conditions of darkness; the material was then ground. The plant was botanically identified by Abigail Aguilar, M.Sc., and a voucher specimen was deposited at the Herbarium of the Instituto Mexicano del Seguro Social, Mexico (IMSSM) with code number 14256.

Preparation of extracts

The dry and powdered plant material (1.364 kg) was macerated three times with CHCl₃ analytical reagent - AR- (J.T. Baker) at room temperature for 7 days. The extract was filtered and concentrated to dryness under low pressure at 40°C. The plant material was later macerated with EtOH AR grade (J.T. Baker) three times for 7 days and the solvent was eliminated under reduced pressure until the solvent-free extract was obtained.

Antiprotozoal activity evaluation

For this assay, the E. histolytica strain HM1-IMSS and the T. vaginalis GT9 strain were cultured in a TYI-S-33 modified medium supplemented with 10% calf serum; G. lamblia strain IMSS:0989:1 was maintained in a TYI-S-33 medium supplemented with 10% calf serum and bovine bile. In vitro susceptibility assays were performed according to the method previously described [25,26]. Briefly, 5×10^4 trophozoites of *G. lamblia* were incubated for 48 h at 37°C with increasing concentrations of the EtOH and CHCl3 extracts of P. americana seeds using Dimethyl sulfoxide (DMSO) as a suitable solvent. After incubation, G. lamblia trophozoites were washed and subcultured for an additional 48 h in a fresh medium alone. For E. histolytica and T. vaginalis, 6 × 10³ trophozoites were incubated for 72 h at 37°C with increasing concentrations of the samples tested. Metronidazole was included as a positive control; parasites without treatment but with the highest DMSO concentration used for sample dilutions were included as a negative control. G. lamblia, T. vaginalis and E. histolytica trophozoites were counted and the 50% Inhibitory concentration (IC₅₀) was calculated by Probit analysis. The experiments were carried out in triplicate and repeated at least twice.

Antimycobacterial evaluation

For this assay, 12 Mycobacterium strains were employed: M. tuberculosis H37Rv (ATCC 27294, a strain sensitive to streptomycin (STR), isoniazid (INH), rifampicin (RIF), etambutol (EMB), or pyrazinamide; M. tuberculosis SIN 4 (a MDR clinical isolate with resistance to first-line drugs); M. tuberculosis MMDO (an MDR clinical isolate with resistance to INH and EMB); four mono-resistant strains of *M. tuberculosis* H37Rv (INH-R, ATCC 35822; STR-R, ATCC 35820; RIF-R, ATCC 35838, and EMB-R, ATCC 35837) and four NTM clinical isolates (M. fortuitum, M. chelonae, M. abscessus and M. avium) and M. smegmatis (ATCC 35798). The Mycobacterium strains were grown and maintained in Middlebrook 7H9 broth supplemented with 10% OADC enrichment (Becton Dickenson, USA) at 37°C until a logarithmic growth phase was achieved. At the moment of evaluation, M. tuberculosis strains and NTM were diluted 1:20 and 1:50, respectively in the 7H9 medium.

The antimycobacterial activity was carried out by means of the Microplate alamar blue assay (MABA) previously described [26,27]. EtOH and CHCl₃ extracts (10 mg) were solubilized in 500 μ L of DMSO and from these stock solutions, several dilutions were prepared to achieve concentrations between 200 and 3.13 μ g/ml. The maximum DMSO concentration used in the MABA assay does not affect mycobacterial growth, as it was previously reported [28]. Extracts that presented MIC <100 μ g/ml were considered to have good antimycobacterial activity. Rifampicin and isoniazid at 0.06 μ g/mL were included as a positive control for the H37Rv strain, for the MDR clinical isolates and for NTM; the same drugs were used but at concentrations of 100 and 3.1 μ g/ml, respectively.

Results and discussion

By means of the maceration process, 88.7 g of the CHCl₃ extract and 77.2 g of the EtOH extract from *P. americana* seeds were obtained with an average yield of 6% with respect to the plant material's dry weight. A preliminary phytochemical analysis by Thin layer chromatography (TLC) of the CHCl₃ extract led to the detection of the presence of β -sitosterol, phytol and palmitic acid. On the other hand, catechin and epicatechin were detected in the EtOH extract by TLC. All compounds were identified by comparison of the Retention factor (R_f) with their commercial reference. A previous study by Rodríguez-Carpena et al. [9] reported a high concentration of catechin and epicatechin in 100% EtOAc, 70% acetone and 70% MeOH extracts obtained from *P. americana* seeds and peel.

The EtOH and CHCl₃ extracts displayed significant antiprotozoal activity. For *G. lamblia, E. histolytica* and *T. vaginalis*, the CHCl₃ extract showed values of $IC_{50} = 0.634$, 0.417 and 0.524 µg/ml, respectively. On the other hand, the EtOH extract exhibited IC_{50}

Table 1 Antiprotozoal activity of the CHCl₃ and EtOH extracts from *Persea americana* seeds

Sample	IC₅₀ (μg/ml)		
	G. lamblia	E. histolytica	T. vaginalis
CHCl ₃ extract	0.634	0.417	0.524
EtOH extract	0.486	0.386	0.533
Metronidazole	0.210	0.06	0.037
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IC₅₀: medium Inhibitory concentration.

values of 0.486, 0.386 and 0.533 μ g/ml against *G. lamblia*, *E. histolytica* and *T. vaginalis*, respectively (Table 1). Although the giardicidal activity of the aqueous seed extract has been previously evaluated [21], the authors used the 3-(4,5-dimetiltiazol-2-ilo)-2,5-difeniltetrazol (MTT) colorimetric method; therefore our data can not be compared.

It is interesting to note that, the reference drug, metronidazole (IC₅₀ = 0.210 μ g/ml) showed only three and two times higher anti-*Giardia* activity than the CHCl₃ and EtOH extracts. However, in the case of *E. histolytica*, the CHCl₃ and EtOH extracts were seven and six times less potent than metronidazole (IC₅₀ = 0.060 μ g/ml). For

Table 2 Antimycobacterial effect of the CHCl₃ and EtOH extracts from *Persea americana* seeds

Mycobacterium tuberculosis	Drug-resistance pattern	MIC (µg/ml)	
		CHCl₃ extract	EtOH extract
H37Rv	INH-, RIF-, STR-, and EMB- susceptible	50	>100
Clinical isolates			
SIN4	STR, INH, RIF, EMB, RFB, ETH, and OFX	50	>100
MMDO	INH, EMB	100	>100
Mono-resistant			
RIF-R	RIF	>100	>100
INH-R	INH	50	>100
STR-R	STR	50	50
EMB-R	EMB	50	50
NTB Mycobacterium			
M. fortuitum		50	100
M. avium		25	100
M. chelonae		100	100
M. smegmatis		12.5	25
M. abscessus		25	100

SIN 4 MDR clinical isolate of *M. tuberculosis* resistant to STR: Streptomycin, INH: Isoniazid, RIF: Rifampicin, EMB: Ethambutol, RFB: Rifabutin, ETH: Ethionamide and OFX: Ofloxacin. MMDO MDR clinical isolates of *M. tuberculosis* resistant to INH and EMB; the resistance pattern was determined by Microdilution alamar blue assay (MABA). NTB: non-tuberculosis mycobacteria; MIC: Minimum inhibitory concentration. Data are means of three determinations. *T. vaginalis*, these extracts showed weak activity, being 16 (CHCl₃) and 17 (EtOH) times less potent than metronidazole (CI₅₀ = $0.037 \ \mu g/ml$).

In traditional Mexican medicine, *P. americana* seeds are used to treat diarrhea [1-4]. It is shown here that *P. americana* EtOH and CHCl₃ seed extracts are indeed responsible for the activity against two anaerobic parasites that cause diarrhea: *G. lamblia*, and *E. histolytica*. The *in vitro* and *in vivo* giardicidal activity of epicatechin isolated from plants such as *Rubus coriifolius* and *Heliantemum glomeratus* has been reported [29,30]. In this regard, epicatechin was detected in *P. americana* EtOH seed extract by TLC analysis; hence, this compound may be responsible for the giardicidal activity observed in this study. Further studies are required to define the active compound(s) responsible for the antiprotozoal activity of the CHCl₃ extract.

It is worth considering the fact that the EtOH and $CHCl_3$ extracts constitute potential sources of compounds that can be employed as prototype molecules for the development of novel antiprotozoal agents as an alternative treatment of clinical isolates with metronidazole resistance.

The results of the antimycobacterial activity of the EtOH and CHCl₃ extracts determined by MABA assay are presented in Table 2. It is important to note that the CHCl₃ extract inhibited the growth of *M. tuberculosis* H37Rv, MDR *M. tuberculosis* SIN 4 and three out of four mono-resistant reference strains of *M. tuberculosis* H37Rv (INH-R, STR-R, and EMB-R), showing a MIC = 50 µg/ml. This extract was also active against the NTM: *M. fortuitum, M. avium, M. smegmatis* and *M. abscessus* with MIC values < 50 µg/ml. However, the EtOH extract affected only the growth of *M. smegmatis* (MIC = 25 µg/ml) and the mono-resistant strains of *M. tuberculosis* H37Rv STR-R and EMB-R (MIC = 50 µg/ml).

To date, the hexanic and MeOH extracts obtained from the stems and leaves of *P. americana* have been reported to inhibit the growth of *M. tuberculosis* H37Rv and *M. tuberculosis* H37Ra strains [18,31]. The reported antimycobacterial activity of the MeOH extracts was attributed to the presence of lignans [31] and the MIC was slightly lower than the CHCl₃ extract tested in this study.

Since the presence of MDR and extensively drugresistant (XDR) *M. tuberculosis* cases is increasing rapidly and current chemotherapy is prolonged, poorly effective, expensive and is accompanied by severe side effects [26]; it is necessary to have recourse to an alternative treatment against these strains or that can even aid in and/or shorten the currently available that could have a different mechanism of action. Therefore, the high activity exhibited by the *P. americana* CHCl₃ seed extract against mono-resistant strains of *M. tuberculosis* H37Rv, MDR clinical isolates and NTM that is described in this study, it is of great interest.

From the clinical point of view, NTM are becoming relevant, because of the so-called mycobacteriosis and are currently recognized as important pathogens associated with both immune-deficient and immune-competent patients. The mycobacteria tested in this study are representative of the most common NTM isolated from pulmonary cases (M. abscessus and M. avium) or associated with soft tissue infections (M. fortuitum and M. chelonae) [32]. Although *M. smegmatis* is a poor pathogenic bacterium, it was included in the NTM group because it is widely employed in the determination of the antimycobacterial activity of new compounds [32,33]. In the case of NTM, the majority is naturally resistant to some of the first-line anti-TB drugs such as INH and RIF; thus, effective drugs against NTM are scarce than those for TB, emphasizing the urgency of finding novel active compounds that could be used in the treatment of the NTM group. Based on our results, P. americana seeds may be a source for potential moieties (molecules) against NTM. We are currently conducting the isolation and identification of the active compounds responsible for the antimycobacterial activity observed with the CHCl₃ extract.

Conclusions

Herein, to the best of our knowledge, the activity of the $CHCl_3$ and EtOH seed extracts from *P. americana* against two intestinal parasites that cause diarrhea: *E. histolytica*, and *G. lamblia* has been demonstrated. In addition, based on our results, $CHCl_3$ seed extract may be a source for potential moieties (molecules) against *M. tuberculosis* drug-resistant species as well as NTM.

Further studies are required for the identification of the active compounds responsible for the antiprotozoal and antimycobacterial activity observed with the EtOH and $CHCl_3$ seed extracts from *P. americana*.

Abbreviations

ALT: Alanine aminotransferase; AST: Aspartate aminotransferase; AOA: Antioxidant activity; CHCl3: Chloroformic; AR: Analytical reagent; DMSO: Dimethyl sulfoxide; EtOAc: Ethyl acetate; EtOH: Ethanol; MeOH: Methanol; MTT: 3-(4,5-dimetiltiazol-2-ilo)-2,5-difeniltetrazol; IMSSM: Instituto Mexicano del Seguro Social Mexico; P. americana: Persea americana; HDL: High density lipoprotein; LDL: Low density lipoprotein; R_f: Retention factor; TLC: Thin layer chromatography; EMB: Etambutol; INH: Isoniazid; STR: Streptomycin; RIF: Rifampicin; LC₅₀: Lethal concentration; LD₅₀: Lethal dose; MIC: Minimum inhibitory concentration; IC₅₀: Medium Inhibitory concentration; MABA: Microplate alamar blue assay; MDR: Multidrug-resistant; XDR: Extensively drug-resistant; M. fortuitum: Mycobacterium fotuitum; M. avium: Mycobacterium avium; M. smegmatis: Mycobacterium smegmatis; M. absessus: Mycobacterium absessus; M. tuberculosis: Mycobacterium tuberculosis; NTM: non-tuberculosis mycobacterium: F. histolytica: Entamoeba histolytica: G. lamblia: Giardia lamblia; T. vaginalis: Trichomonas vaginalis.

Competing interests

The authors declare that they have no competing interests.

Authors' contributions

AJ-A designed and coordinated the study, prepared the $CHCI_3$ and EtOH extracts and carried out their phytochemical analysis and wrote the

manuscript. C-G collected the *P. americana* seeds and contributed to the preparing the CHCl₃ and EtOH extracts. JL-H and RR-N evaluated the antimycobacterial activity from CHCl₃ and EtOH extracts. The antiprotozoal activity from both extracts was determined by AT and LY-M, who also contributed to the manuscript preparation. All authors have read and approved the final manuscript.

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References

- Aguilar A, Aguilar A: Herbario Medicinal del IMSS. 1st edition. México: Editorial Redact, S.A; 1994.
- Argueta A, Cano L, Rodarte M: Atlas de las Plantas de la Medicina Tradicional Mexicana, Vols. 2, 3. 1st edition. México, D.F: Editorial Instituto Nacional Indigenista; 1994.
- Moreno-Uribe V: Herbolaria y tradición en la región de Xico. 1st edition. Mexico: Veracruz, Diseño Editorial; 2008.
- Osuna-Torres L, Tapia-Pérez ME, Aguilar-Contreras A: Plantas medicinales de la medicina tradicional mexicana para tratar afecciones gastrointestinales. 1st edition. España: Editorial Universidad de Barcelona; 2005.
- Del Refugio Ramos M, Jerz G, Villanueva S, López-Dellamary F, Waibel R, Winterhalter P: Two glucosylated abscicic acid derivates from avocado seeds (*Persea americana* Mill. Lauraceae cv. Hass). *Phytochem* 2004, 65(7):955–962.
- Nwaogu LA, Alisi CS, Ojiako OA: Studies on the nutritional and phytochemical properties of *Persea americana* seed. *Bio-Research* 2008, 6(1):320–322.
- Takenaga F, Matsuyama K, Abe S, Torii Y, Itoh S: Lipid and fatty acid composition of mesocarp and seed of avocado fruits harvested at northern range in Japan. J Oleo Sci 2008, 57(11):591–597.
- Wang W, Bostic TR, Gu L: Antioxidant capacities, procyanidins and pigments in avocados of different strains and cultivars. *Food Chem* 2010, 122(4):1193–1198.
- Rodríguez-Carpena JG, Morcuende D, Andrade MJ, Kylli P, Estévez M: Avocado (*Persea americana* Mill.) phenolics, *in vitro* antioxidant and antimicrobial activities, and inhibition of lipid and protein oxidation in porcine patties. J Agric Food Chem 2011, 59(10):5625–5635.
- Asaolu MF, Asaolu SS, Oyeyemi AO, Aluko BT: Hypolipemic effects of methanolic extract of *Persea americana* seeds in hypercholesterolemic rats. J Med Medical Sciences 2010, 1(4):126–128.
- Imafidon KE, Amaechina FC: Effects of aqueous seed extract of *Persea* americana Mill. (avocado) on blood pressure and lipid profile in hypertensive rats. *Adv Biol Res* 2010, 4(2):116–121.
- Ogochukwu NA, Ozolua RI, Okpo SO: Effect of the aqueous seed extract of Persea americana Mill. (Lauraceae) on the blood pressure of Sprague Dawley rats. Afric J Pharm Pharmacol 2009, 3(10):485–490.
- Edem D, Ekanem I, Ebong P: Effect of aqueous extracts of alligator pear seed (*Persea americana* Mill.) on blood glucose and histopathology of pancreas in alloxan-induced diabetic rats. *Pak J Pharm Sci* 2009, 22(3):272–276.
- Koffi N, Kouakou EA, Dodiomon S: Effect of aqueous extract of Persea americana seeds on the glycemia of diabetic rabbits. Europ J Sci Res 2009, 26(3):376–385.
- Okonta M, Okonta L, Aguwa CN: Blood glucose lowering activities of seeds of *Persea americana* on alloxan induced diabetic rats. *Nig J Nat Prod Med* 2007, 11:26–28.

- Ozolua RI, Anaka ON, Okpo SO, Idogun SE: Acute and sub-acute toxicological assessment of the aqueous seed extract of *Persea americana* Mill (Lauraceae) in rats. *Afr J Tradit Complement Altern Med* 2009, 6(4):573–578.
- Giffoni LJ, Salles EH, Aguiar R, Nogueira RS, Costa JJ, Medeiros SL, De Morais S, Gadelha MF: Chemical composition, toxicity and larvicity and antifungal activities of *Persea americana* (avocado) seed extracts. *Rev Soc Bras Med Trop* 2009, 42:110–113.
- Gomez-Flores R, Arzate-Quintana C, Quintanilla-Licea R, Tamez-Guerra P, Tamez-Guerra R, Monreal-Cuevas E, Rodríguez-Padilla C: Antimicrobial activity of *Persea americana* Mill (Lauraceae) (Avocado) and *Gymnosperma glutinosum* (Spreng.) Less (Asteraceae) leaf extracts and activities fractions against *Mycobacterium tuberculosis*. *Amer-Eur J Scient Res* 2008, 3(2):188–194.
- Raymond Chia TW, Dykes GA: Antimicrobial activity of crude epicarp and seed extracts from mature avocado fruit (*Persea americana*) of three cultivars. *Pharm Biol* 2011, 48(7):753–756.
- Abe F, Nagafuji S, Okawa M, Kinjo J, Akahane H, Ogura T, Martínez-Alfaro MA, Reyes-Chilpa R: Trypanocidal constituents in plants. 5. Evaluation of some Mexican plants for their trypanocidal activity and active constituents in the seed of *Persea americana*. *Biol Pharm Bull* 2005, 28(7):1314–1317.
- Ponce-Macotela M, Navarro-Alegría I, Martínez-Gordillo MN, Álvarez-Chacón R: Efecto antigiardiásico in vitro de 14 extractos de plantas. *Rev Invest Clin* 1994, 46:343–3474.
- Matsusaka Y, Kawabata J, Takanori K: Antioxidative constituents in avocado (*Persea americana* Mill.) seeds. J Jap Soc Food Sci Technol 2003, 50(11):550–552.
- 23. Yean-Yean S, Barlow PJ: Antioxidant activity and phenolic content of selected fruit seeds. *Food Chem* 2004, 88(3):411–417.
- Asaolu MF, Asaolu SS, Fakunle JB, Emman-Okon BO, Ajayi EO, Togun RA: Evaluation of *in vitro* antioxidant activities of methanol extracts of *Persea americana* and *Cnidosculus aconitifolius*. *Pak J Nutr* 2010, **9:**1074–1077.
- Cedillo-Rivera R, Chávez B, González-Robles A, Tapia A, Yépez-Mulia L: In vitro effect of nitazoxanide against Entamoeba histolytica, Giardia intestinalis and Trichomonas vaginalis trophozoites. J Eukaryot Microbiol 2002, 49(3):201–208.
- Jiménez-Arellanes A, León-Díaz R, Meckes M, Tapia A, Molina-Salinas GM, Luna-Herrera J, Yépez-Mulia L: Antiprotozoal and antimycobacterial activities of pure compounds from Aristolochia elegans rizhomes. Evid Based Complement Alternat Med 2012, 201(2):593403. doi:10.1155/2012/593403.
- León-Díaz R, Meckes M, Said-Fernández S, Molina-Salinas GM, Vargas-Villareal J, Torres J, Luna-Herrera J, Jiménez-Arellanes A: Antimycobacterial neolignans isolated from Aristolochia taliscana. Mem Inst Oswaldo Cruz 2010, 105(1):45–51.
- Molina-Salinas GM, Ramos-Guerra MC, Vargas-Villarreal J, Mata-Cárdenas BD, Becerril-Montes P, Said-Fernández S: Bactericidal activity of organic extracts from *Fluorensia cernua* DC against strains of *Mycobacterium tuberculosis*. Arch Med Res 2006, 37(1):45–49.
- Alanis AD, Calzada F, Cedillo-Rivera R, Meckes M: Antiprotozoal activity of the constituents of Rubus coriifolius. *Phytother Res* 2003, 17(6):681–682.
- Barbosa E, Calzada F, Campos R: *In vivo* antigiardial activity of three flavonoids isolated of some medicinal plants used in Mexican traditional medicine for the treatment of diarrhea. *J Ethnopharmacol* 2007, 109(3):552–554.
- Baquero SE, Benavides J, Sepúlveda L, Quiñones W, Torres F, Cardona D, Archbold R, Guzmán JD, Cuca LE, Franzblau S, Echeverri F: Actividad antimicobacteriana de algunas plantas de la flora colombiana. *Sci Techn* 2007, XIII:133–136.
- Cassidy PM, Hedberg K, Saulson A, McNelly E, Winthrop KL: Nontuberculous mycobacterial disease prevalence and risk factors: a changing epidemiology. *Clin Infect Dis* 2009, 49(12):e124–e129.
- Pauli GF, Case RJ, Inui T, Wang Y, Cho S, Fischer NH, Franzblau SG: New perspectives on natural products in TB drug research. *Life Sci* 2005, 78(5):485–494.

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